## Amendments to the Specification:

Please amend Paragraph [0001] as follows:

[0001]

This is a division of co-pending application Serial No. 09/738,363 (filed December 15, 2000) which is a division of application Serial No. 09/076,137 (filed on May 12, 1998, which issued as U.S. Patent No. 6,166,195 on December 26, 2000) which is a division of application Serial No. 08/316,301 (filed on September 30, 1994, which issued as U.S. Patent No. 5,753,492 on May 19, 1998) which is a division of application Serial No. 07/871,510 (filed on April 23, 1992, now abandoned) which is a continuation-in-part of application Serial No. 07/693,018 (filed on May 3, 1991, now abandoned) and a continuation-in-part of application Serial No. 07/830,050 (filed on January 31, 1992, now abandoned). Serial No. 07/693,018 was a continuation-in-part of Serial No. 07/565,544 (filed on August 10, 1990, now abandoned) which is a continuation-inpart of application Serial No. 07/084,653 (filed on August 12, 1987, now U.S. Patent No. 4,948,734). The subject application is also a continuation-in-part of application Serial No. 07/675,772 (filed March 27, 1991, now U.S. Patent No. 5,262,399) which is a continuation-inpart of Serial No. 07/565,544 and a continuation in part of Serial No. 07/557,246 (filed on July 24, 1990, now U.S. Patent No. 5,281,530). Serial No. 07/557,246 is a continuation-in-part of Serial No. 07/535,810 (filed June 11, 1990, now abandoned) which is a continuation-in-part of Serial No. 07/084,653.

Please amend Paragraph [0129] as follows:

[00129]

RFLP analyses were performed by standard hybridization of Southern blots of PS52A1 DNA with a <sup>32</sup>P-labeled oligonucleotide probe designed from the N-terminal amino acid sequence disclosed in Example 2. The sequence of this probe is:

5' ATG ATT ATT GAT TCT AAA ACA ACA TTA CCA AGA CAT TCA/T TTA ATA/T AAT ACA/T ATA/T AA 3' (SEQ ID NO. 25)

This probe was designated 52A1-C. Hybridizing bands included an approximately 3.6 kbp *HindIII* fragment and an approximately 8.6 kbp *EcoRV* fragment. A gene library was

constructed from PS52A1 DNA partially digested with Sau3A. Partial restriction digests were fractionated by agarose gel electrophoresis. DNA fragments 6.6 to 23 kbp in size were excised from the gel, electroeluted from the gel slice, and recovered by ethanol precipitation after purification on an Elutip-D ion exchange column. The Sau3A inserts were ligated into BamHIdigested LambdaGem-11 (Promega). Recombinant phage were packaged and plated on E. coli KW251 cells (Promega). Plaques were screened by hybridization with the radiolabeled 52A1-C oligonucleotide probe disclosed above. Hybridizing phage were plaque-purified and used to infect liquid cultures of E. coli KW251 cells for isolation of phage DNA by standard procedures (1x SSC and 0.1% SDS at 68°C) (Maniatis et al.). For subcloning, preparative amounts of DNA were digested with EcoRI and SalI, and electrophoresed on an agarose gel. The approximately 3.1 kbp band containing the toxin gene was excised from the gel, electroeluted from the gel slice, and purified by ion exchange chromatography as above. The purified DNA insert was ligated into EcoRI + SalI-digested pHTBlueII (an E. coli/B. thuringiensis shuttle vector comprised of pBluescript S/K [Stratagene] and the replication origin from a resident B.t. plasmid [D. Lereclus et al. 1989. FEMS Microbiology Letters 60:211-218]). The ligation mix was used to transform frozen, competent E. coli NM522 cells (ATCC 47000). Transformants were plated on LB agar containing ampicillin, isopropyl-(Beta)-D-thiogalactoside (IPTG), and 5-Bromo-4-Chloro-3indolyl-(Beta)-D-galactoside (XGAL). Plasmids were purified from putative recombinants by alkaline lysis (Maniatis et al.) and analyzed by electrophoresis of EcoRI and SalI digests on agarose gels. The desired plasmid construct, pMYC2321 contains a toxin gene that is novel compared to the maps of other toxin genes encoding nematicidal proteins.